



Complete Genome Sequence of *Serratia marcescens* SmUNAM836, a Nonpigmented Multidrug-Resistant Strain Isolated from a Mexican Patient with Obstructive Pulmonary Disease

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***Serratia marcescens* SmUNAM836 is a multidrug-resistant clinical strain isolated in Mexico City from a patient with chronic obstructive pulmonary disease. Its complete genome sequence was determined using PacBio RS II SMRT technology, consisting of a 5.2-Mb chromosome and a 26.3-kb plasmid, encoding multiple resistance determinants and virulence factors.**

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Serratia marcescens is emerging as an opportunistic nosocomial pathogen causing urinary tract, bloodstream, skin, and respiratory infections (1). Several virulence factors have been identified, such as hemolysins, proteases, and siderophores (2). Many clinical isolates of this species display intrinsic or acquired resistance to multiple antibiotic families (1).

Here we present the complete genome sequence of *Serratia marcescens* strain SmUNAM836, a nonpigmented, multidrug-resistant bacterium isolated from the bronchial aspiration of a hospitalized patient with chronic obstructive pulmonary disease in Mexico City, as detailed in BioProject accession no. PRJNA284857. Strain SmUNAM836 was resistant to diverse β -lactams, including penicillins, but it is susceptible to cephalosporins and carbapenems. It is resistant to aminoglycosides (tobramycin), fluoroquinolones (ciprofloxacin), and polymixin B.

The genomic DNA of the strain was purified with the DNeasy blood and tissue kit (Qiagen) and sent to the Yale Center for Genome Analysis (YCGA) for PacBio RS II single-molecule real-time (SMRT) sequencing. A standard library of 10-kb fragments was prepared and sequenced on three SMRT cells with the P2-C4 chemistry. The continuous long reads were assembled using the HGAP/Quiver protocol in SMRT portal v.2.3.0.140936.p4 (3), resulting in an assembly with 2 contigs. They were circularized by trimming the terminal repeats with Minimus2 (4) and subjected to two consecutive rounds of read remapping with the RS_Resequencing.1 module for sequence polishing. The final assembly had a mean coverage of \sim 172 \times and consists of a chromosome (5,207,023 bp), and a plasmid (26,346 bp), with a mean G+C content of 59.71%. Gene calling and annotation were performed with a modified version of Prokka (5). A total of 4,961 genes were annotated: 4788 CDSs, 95 tRNAs, 22 rRNAs, and 1 tmRNA. Additionally, 45 ncRNAs, 3 riboswitches, and 502 signal peptides were also identified, but no clustered regularly interspaced short palindromic repeat (CRISPR) arrays. The annotation was manually curated and enriched by adding 6 prophage predictions made

by the PHAST server (6) and 13 genomic islands identified by IslandViewer3 (7).

The genome was screened for the presence of antibiotic resistance genes, efflux pumps, and potential virulence factors using blastp (8) searches against locally maintained versions of the CARD (9), ResFinder (10), and VirulenceFinder (11) databases. Eight resistance-nodulation-division (RND), two ATP-binding cassette (ABC), and one major-facilitator-superfamily (MFS) type efflux pumps were identified, along with the chromosomally encoded BlaKPC-2 (class B) and AmpC-like (class C) beta-lactamases, the aminoglycoside acetyltransferase Aac(6')-Ic, and the aminoglycoside phosphotransferase StrA, an QnrB31-like fluoroquinolone resistance protein. No class 1 or class 2 integrons or ISCR elements were found.

The plasmid had a low G+C content (43.52%) and displayed \sim 97% sequence identity with diverse *Enterobacter cloacae* plasmids like pKPC-f91. It harbors the transposon Tn3 encoding 3 proteins: a BlaTEM-1D (class A) beta-lactamase and the Tn3 transposase and resolvase. It encodes the genes for the *cdAB* toxin-antitoxin system.

Potential virulence factors detected are long polar fimbriae (LpfA), hemolysin D (HlyD), and the ClpP protease, the IroN enterobactin siderophore receptor, hemolytic phospholipase C (PlcH), and plasmid-encoded catalase peroxidase (KatP) and enterotoxin (SenB).

Nucleotide sequence accession numbers. The complete genome sequence of *Serratia marcescens* SmUNAM836 is available in GenBank under accession numbers **CP012685** (chromosome) and **CP012686** (plasmid), BioProject accession number PRJNA284857, and BioSample accession number SAMN03733572.

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REFERENCES

- Iguchi A, Nagaya Y, Pradel E, Ooka T, Ogura Y, Katsura K, Kurokawa K, Oshima K, Hattori M, Parkhill J, Sebaihia M, Coulthurst SJ, Gotoh N, Thomson NR, Ewbank JJ, Hayashi T. 2014. Genome evolution and plasticity of *Serratia marcescens*, an important multidrug-resistant nosocomial pathogen. *Genome Biol Evol* 6:2096–2110.
- Shimuta K, Ohnishi M, Iyoda S, Gotoh N, Koizumi N, Watanabe H. 2009. The hemolytic and cytolytic activities of *Serratia marcescens* phospholipase A (PhlA) depend on lysophospholipid production by PhlA. *BMC Microbiol* 9:261.
- Chin C, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. *Nat Methods* 10:563–569. <http://dx.doi.org/10.1038/nmeth.2474>.
- Treangen TJ, Sommer DD, Angly FE, Koren S, Pop M. 2012. Next generation sequence assembly with AMOS. *Curr Protoc Bioinformatics* Chapter 11:Unit 11:8. <http://dx.doi.org/10.1002/0471250953.bi1108s33>.
- Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 30:2068–2069. <http://dx.doi.org/10.1093/bioinformatics/btu153>.
- Zhou Y, Liang Y, Lynch KH, Dennis JJ, Wishart DS. 2011. PHAST: a fast phage search tool. *Nucleic Acids Res* 39:W347–W352. <http://dx.doi.org/10.1093/nar/gkr485>.
- Dhillon BK, Laird MR, Shay JA, Winsor GL, Lo R, Nizam F, Pereira SK, Waglechner N, McArthur AG, Langille MGL, Brinkman FSL. 2015. IslandViewer 3: More flexible, interactive genomic island discovery, visualization and analysis. *Nucleic Acids Res* 43(Web Server Issue): W104–W108. <http://dx.doi.org/10.1093/nar/gkv401>.
- Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+: architecture and applications. *BMC Bioinformatics* 10:421. <http://dx.doi.org/10.1186/1471-2105-10-421>.
- McArthur AG, Waglechner N, Nizam F, Yan A, Azad MA, Baylay AJ, Bhullar K, Canova MJ, De Pascale G, Ejim L, Kalan L, King AM, Koteva K, Morar M, Mulvey MR, O'Brien JS, Pawlowski AC, Piddock LJ, Spanogiannopoulos P, Sutherland AD, Tang I, Taylor PL, Thaker M, Wang YM, Yu T, Wright GD. 2013. The comprehensive antibiotic resistance database. *Antimicrob Agents Chemother* 57:3348–3357. <http://dx.doi.org/10.1128/AAC.00419-13>.
- Zankari E, Hasman H, Cosentino S, Vestergaard M, Rasmussen S, Lund O, Aarestrup FM, Larsen MV. 2012. Identification of acquired antimicrobial resistance genes. *J Antimicrob Chemother* 67:2640–2644. <http://dx.doi.org/10.1093/jac/dks261>.
- Kleinheinz KA, Joensen KG, Larsen MV. 2014. Applying the ResFinder and VirulenceFinder web-services for easy identification of acquired antibiotic resistance and virulence genes in bacteriophage and prophage nucleotide sequences. *Bacteriophage* 4:e27943. <http://dx.doi.org/10.4161/bact.27943>.